



---

## ROLE OF BIOFERTILIZERS IN INTEGRATED PLANT NUTRIENT SYSTEM (IPNS)

**Dr. Divya Vyas**

Senior Research Fellow, Department of Agriculture chemistry and Soil Science, Rajasthan College of Agriculture, Maharana Pratap University of Agriculture and Technology, Udaipur, Rajasthan -313001.

**Dr. Ram Hari Meena**

Assitant Professor, Department of Agriculture chemistry and Soil Science, Rajasthan College of Agriculture, Maharana Pratap University of Agriculture and Technology, Udaipur, Rajasthan - 313001.

### ABSTRACT

*In India, framers doing intensive agriculture which led to rapid depletion of nutrient in agricultural land which is balanced by use of chemical based fertilizers. Chemical fertilizers not only disturb ecological balance but also pose health hazards besides these are quite expensive, bringing the cost of production very high. Farmers use to indulge money in purchasing costly fertilizers. So been an agriculturist we have to focus on alternative of fertilization which maintain ecological balance as well as reduce farmers expenses. Biofertilizers are low cost and eco-friendly inputs and have tremendous potential for supplying nutrients, especially N and P, and can reduce the chemical fertilizer dose by 25-50%. This review helps towards use of biofertilizers in sustainable agriculture and reducing problems associated with the use of chemicals fertilizers.*

**Keywords:** Biofertilizer, Chemical fertilizer, Integrated nutrient management, Sustainable agriculture

## Introduction

In the last century, with the concept of intensive agriculture the use chemical fertilizers is more and this made farmers to be happy of getting increased yield in agriculture in the beginning. But slowly chemical fertilizer started displaying their ill effects such as leaching, polluting water basins, destroying microorganisms and friendly insects, making the crop more susceptible to the attack of diseases, reducing the soil fertility, pose health hazards besides these are quite expensive, bringing the cost of production much higher and thus causing irreparable damage to the overall system. Chemical fertilizers are industrially manipulated, substances composed of known quantities of nitrogen, phosphorus and potassium, and their exploitation causes air and ground water pollution by eutrophication of water bodies (Youssef *et al*; 2014). In general, 60% to 90% of the total applied fertilizer is lost and the remaining 10% to 40% is taken up by plants. The results of long-term fertility experiments have also clearly demonstrated that chemical fertilizers alone cannot sustain the productivity at current level under intensive cropping systems (Swarup *et al.* 1998).

International Plant Nutrition Institute has confirmed that Indian Soils are under nutrient crisis and also concluded that in the absence of nutrient balance sheet of Indian Agriculture will continue to be negative (TNAU, 2009). This necessitated the scope to find out suitable alternative nutrient combinations for different crops to overcome the burden of chemical fertilizers and for achieving the higher productivity. Currently, the biological approaches for improving crop production are gaining strong status among agronomists and environmentalists following integrated plant nutrient management system ( Ahemad and Kibret, 2014). So, there is urgent need to encourage alternate means of soil fertilization relies on organic inputs to improve nutrient supply and conserve the field management (Araujo *et al.*, 2008). Minerals, organic components and microorganisms are three major solid components of the soil. They profoundly affect the physical, chemical, and biological properties and processes of terrestrial systems. Organic manures are helpful to improve the physical and chemical properties which results into increasing soil fertility and productivity (Patil *et al.*, 2013). Organic manure greatly reduces leaching of fertilizer, pesticide and herbicides into the ground water. Organic manure improves the activity of earthworm and other soil micro flora and it increases soil infiltration rate and reduces soil evaporation there by it increases soil water storage. The organic manure influences

agricultural sustainability by enhancing productivity (Singh et al., 2016). Another important component of soil is beneficial microorganism which are abundant in soil as well as in biofertilizers. A major focus in the coming decades would be on safe and eco-friendly methods by exploiting the beneficial micro-organisms in sustainable crop production (Nina, 2014). Biofertilizers are low cost and eco-friendly inputs and have tremendous potential for supplying nutrients, especially N and P, and can reduce the chemical fertilizer dose by 25-50% (Vance 1997; Rana et al., 2012).

According to the definition proposed by Vessey [2003], biofertilizers are substances which contain living microorganisms which, when applied to seed, plant surfaces, or soil, colonize the rhizosphere or the interior of the plant, and promote growth by increasing the supply or availability of primary nutrients to the host plant. Biofertilizers keep the soil environment rich in all kinds of micro- and macro-nutrients via nitrogen fixation, phosphate and potassium solubalisation or mineralization, release of plant growth regulating substances, production of antibiotics and biodegradation of organic matter in the soil (Sinha, 2014). Through the use of bio-fertilizers, healthy plants can be grown, while enhancing the sustainability and the health of the soil. It can also provide protection against drought and some soil-borne diseases. It act as eco-friendly and cost effective inputs for the farmers ((Nilabja Ghosh, 2004, Mohammadi, K., and Yousef, S. 2012). The role and importance of biofertilizers in sustainable crop production has been reviewed by several authors (Biswas *et al.* 1985; Wani and Lee, 1995; Katyayal *et al.* 1994). Thus, biofertilizers can be important components of integrated nutrients management. Integrate nutrient management plays a key role in modern agriculture in increasing the productivity of crops and sustained management of soil fertility (Singh, G. et al. 2016). They hold vast potential in meeting plant nutrient requirements while minimizing the use of chemical fertilizers. It is important to realise the useful aspects of biofertilizers and implement its application to modern agricultural practices. The new technology developed using the powerful tool of molecular biotechnology.

In order to meet the food requirements of ever increasing population, the fertilizer requirement for crop production is very high. Biological nitrogen fixation can be the key to fill up this gap because of high cost and several other demerits of chemical fertilizers. For production of a good and efficient biofertilizer, first of all an efficient strain is required, then its inoculums (the form in which the strain is to be applied in fields) is produced. Several

microorganisms and their association with crop plants are being exploited in the production of biofertilizers. They can be grouped in different ways based on their nature and function **Table-1**. While producing bio-fertilizers the standards laid down by BIS have also to be kept in mind for making the product authentic. Inoculation of *Rhizobium* and application of fertilizer improved the yield attributes grain and straw yield (Balchandran and Nagarajan, 2002). Several reports have indicated that biofertilizers like *Azotobacter* and PSB alone or in combination with chemical fertilizers have great prospect in increasing productivity of wheat (Kumar and Ahlawat 2004).

Table 1: Table showing different groups of microorganism and their example

<b>Groups</b>	<b>Examples</b>
<b>N fixing biofertilizers</b>	
Free living	<i>Azotobacter, Beijerinckia, Clostridium, Klebsiella, Anabaena</i> and <i>Nostoc</i>
Symbiotic	<i>Rhizobium, Frankia</i> and <i>Anabaena azollae</i>
Associative symbiotic	<i>Azospirillum</i>
<b>P solubilising biofertilizers</b>	
Bacteria	<i>Bacillus megaterium, phosphaticum, Bacillus subtilis, B. circulans</i> and <i>Pseudomonas striata</i>
Fungi	<i>Penicillium sp.</i> and <i>Aspergillus awamori</i>
<b>P mobilizing biofertilizers</b>	
Arbuscular mycorrhiza	<i>Glomus sp., Gigaspora sp., Acaulospora sp., Scutellospora sp.</i> and <i>Sclerocystis sp.</i>
Ectomycorrhiza	<i>Laccaria sp., Pisolithus sp., Boletus sp.</i> And <i>Amanita sp.</i>
Ericoid mycorrhiza	<i>Pezizella ericae</i>
Orchid mycorrhiza	<i>Rhizoctonia solani</i>
<b>Biofertilizers for micro nutrients</b>	
Silicate and Zinc solubilizers	<i>Bacillus sp.</i>
<b>Plant growth promoting rhizobacteria</b>	
<i>Pseudomonas</i>	<i>Pseudomonas fluorescens</i>

**N-fixing Biofertilizers (NBF):** Bio-fertilizers are the formulations of living microorganisms which are able to fix atmospheric nitrogen in the available form for plants (nitrate form) either by living freely in the soil or associated symbiotically with plants. Although nitrogen fixers are present in the soil, enrichment of soil with effective microbial strains (**Table 2**) is much beneficial for the crop yields.

Table 2: Some major nitrogen fixing microorganisms and beneficiaries plant

<b>S.No.</b>	<b>Name of microorganisms</b>	<b>Name of crop plants which receive benefits</b>

1	<i>Rhizobium</i> spp. living symbiotically in root nodules	All grain legumes (pulses), some oil yielding (soybean, groundnut), some fodder legumes (rizka and berseem)
2	Nostoc, Anabaena, Aulosira and others (free living blue green algae)	Rice
3	<i>Anabaena azollae</i> living symbiotically with the waterfern	Rice, Azolla spp.
4	<i>Azotobacter chroococcum</i> (free living bacterium)	Rice, maize, cotton and others
5	Frankia spp. (actinomycete) living symbiotically in nonlegume root nodules	Alnus, Casuarina and others
6	Azospirillum spp. (associate symbiont) Bacillus polymyxa, Clostridium spp., Rhodospirillum spp.	Maize, sorghum, pearl-millet, finger millet and others non-specific hosts

### (i) Azotobacter (Family: Azotobacteriaceae)

It is the important and well known free living nitrogen fixing aerobic bacterium, belongs to family Azotobacteriaceae. It is used as a biofertilizer for all non leguminous plants especially wheat, rice, sugarcane, cotton, vegetables, oilseeds such as mustard, linseed and for the millets such as pearl millet, finger millets, and kodomillets etc. The Azotobacter colonizing the roots not only remains on the root surface but also a sizable proportion of it penetrates into the root tissues and lives in harmony with the plants. Azotobacters are present in neutral or alkaline soils and *A. chroococcum* is the most commonly occurring species in arable soils. *A. vinelandii*, *A. beijerinckii*, *A. insignis* and *A. macrocytogenes* are other reported species. The lack of organic matter in the soil is a limiting factor for the proliferation of Azotobacter in the soil. The numbers of *A. chroococcum* in Indian soils rarely exceeds  $10^5$ /g soil due to lack of organic matter and the presence of antagonistic microorganisms in soil. They can fix N up to 25 kg/ha under optimum conditions and increase yield up to 50%. The bacterium produces abundant slime which helps in soil aggregation. They also produces certain substances good for the growth of plants and antibodies that suppress many root pathogens. They improve seed germination and plant growth by producing B-vitamins, NAA, GA and other chemicals (plant hormones) that are inhibitory to certain root pathogens (Mazid et al., 2011f). Azotobacter have been found to produce some antifungal substance which inhibits the growth of some soil fungi like *Aspergillus*, *Fusarium*, *Curvularia*, *Alternaria*, *Helminthosporium*, *Fusarium* etc.

Field trials carried out in different locations have demonstrated that under certain environmental and soil conditions inoculation with azotobacteria has beneficial effects on plant yields (Mazid et al., 2011d). For sugarcane, *Azotobacter indicum* is suitable in acidic soils in which it forms *rhizo bacteriocoenotic* association with roots and application in soil is economical but a large amount of organic C- and Mo is needed for stimulating nitrogenase enzyme activity

during N fixation (Mazid et al., 2012b; Khan et al., 2012a). Bahadur *et al.* 2013 reported similar increases in grain and straw yield of wheat due to *Azotobacter* and PSB application in combination with chemical fertilizers indicating increases in N and P uptake by wheat due to *Azotobacter* and PSB biofertilizers due to N<sub>2</sub> fixation and P solubilization in soil.

## ii Symbiotic: Rhizobium (Family: Rhizobiaceae)

They belong to family Rhizobiaceae, symbiotic in nature, fix nitrogen 50-100 kg/ ha with legumes only. The morphology and physiology of Rhizobium will vary from free-living condition to the bacteroid of nodules. Rhizobium has the ability to fix atmospheric N- in symbiotic association with legumes and certain non legumes like, Parasponia (Saikia and Jain, 2007). The bacteria infect the legume root and form root nodules within which they reduce molecular nitrogen to ammonia which is readily utilized by the plant to produce valuable proteins, vitamins and other nitrogen containing compounds. It has been estimated that 40-250 kg N / ha / year is fixed by different legume crops by the microbial activities of Rhizobium.

Pulse crops have unique properties of nodulation through *Rhizobium* bacteria. In recent years use of Rhizobium culture has been routinely recommended as an input in pulse cultivation. In India about 30 million hectares of land is under pulses cultivation. It is useful for pulse legumes like chickpea, red-gram, pea, lentil, black gram, etc., oil-seed legumes like soybean and groundnut and forage legumes like berseem and lucerne. It colonizes the roots of specific legumes to form tumour like growths called root nodules, which act as factories of ammonia production. They have seven genera and highly specific to form nodule in legumes, referred as cross inoculation group, detailed information was given in **Table 3**. The appropriate strain can increase the crop yield up to 10-35% since N is fixed at 40-200 kg/ha which is able to meet up to 80-90% of N need of the crop (Verma 1993). Also, residual N is beneficial for the next crops grown in the same field. Jain *et al.* (2007) reported that *Rhizobium* along with micronutrients significantly enhanced the P and N uptake as compared to control in mungbean.

Table 3: Cross inoculation group of different rhizobia species.

---

<b>Rhizobium sp.</b>	<b>Cross inoculation groups</b>	<b>Legume types</b>
<i>R. leguminosarum</i>	Pea group	Pisum, Vicia, Lens
<i>R. phaseoli</i>	Bean group	Phaseolus
<i>R. trifoli</i>	Clover group	Trifolium
<i>R. lupini</i>	Lupini group	Lupinus, Orinthopus
<i>R. japonicum</i>	Soyabean group	Glycine
<i>R. meliloti</i>	Alfalfa group	Melilotus, Medicago, Trigonella
<i>Rhizobium sp.</i>	Cowpea group	Vigna, Arachis

### (iii) Associative symbiotic: Azospirillum

*Azospirillum* is an associative symbiotic nitrogen fixing bacteria. The bacteria of Genus *Azospirillum* are N<sub>2</sub> fixing organisms isolated from the root and above ground parts of a variety of crop plants. Five species of *Azospirillum* have been described to date *A. brasilense*, *A. lipoferum*, *A. amazonense*, *A. halopraeferens* and *A. irakense*. It fixes the considerable quantity of nitrogen in the range of 20- 40 kg N/ha in the rhizosphere in non-leguminous plants such as cereals, millets, oilseeds, cotton etc. *Azospirillum lipoferum* and *A. brasilense* are primary inhabitants of soil, the rhizosphere and intercellular spaces of root cortex of graminaceous plants. Apart from nitrogen fixation, growth promoting substance production (IAA), vitamins, nicotinic acid, gibberellins, disease resistance and drought tolerance are some of the additional benefits such as better germination, early emergence and better root development.

### (iv) Azolla Family: Azollaceae

Azolla is a free floating water fern that floats in water and fixes atmospheric nitrogen in association with nitrogen fixing blue green algae *Anabaena azollae* a cyanobacterium. Azolla fronds consist of sporophyte with a floating rhizome and small overlapping bi-lobed leaves and roots. *Azolla* is considered to be a potential biofertilizer for wetland rice and it is known to contribute 40-60 kg N/ha per rice crop. Rice growing areas in South East Asia and other third World countries have recently been evincing increased interest in the use of the symbiotic N<sub>2</sub> fixing water fern *Azolla* either as an alternate nitrogen sources or as a supplement to commercial nitrogen fertilizers. For green manuring, Azolla is sown in the field or in a separate shallow pond. Water is drained off the field and Azolla is incorporated into the soil before transplanting of paddy. Dried inoculum of Azolla is also presoaked in 50 ppm of superphosphate solution for 12 h and inoculated in the paddy field (Kannaiyan, 2002). One kg of

it fixes 40-55 kg N/ha, 15-20 P/ha and 20-25 kg K/ha in a month, thus increasing yield of flooded paddy by 10-20% (Ghosh, 2004).

#### **(v) Blue Green Algae (BGA)**

Most N fixing BGA are filamentous, consisting of chain of vegetative cells including specialized cells called heterocyst which function as micro nodule for synthesis and N fixing machinery. BGA forms symbiotic association capable of fixing N- with fungi, liverworts, ferns and flowering plants, but the most common symbiotic association has been found between a free floating aquatic fern, the Azolla and Anabaena azollae (BGA). These phototropic prokaryotic bacteria are effective only in submerged paddy in presence of bright sunlight by forming a bluish-green algae on standing water and by converting the insoluble P into soluble forms, fixing N to the tune of 2-30 kg/ha thereby raising the crop yield by 10-15% when applied at 10kg/ha/BGA biomass. They also produce indole acetic acid (auxin) and gibberllic acid. They too add growth-promoting substances including vitamin B12, improve the soil's aeration and water holding capacity and add to bio mass when decomposed after life cycle.

#### **Phosphate Solubilizing Microorganisms**

Phosphorus (P), the second important plant growth limiting nutrient after nitrogen, is abundantly available in soils in both organic and inorganic forms (Khan et al., 2009). Despite of large reservoir of P, the amount of available forms to plants is generally low (Ezawa et al., 2002). This low availability of phosphorous to plants is because the majority of soil P is found in insoluble forms, while the plants absorb it only in two soluble forms, the monobasic ( $\text{H}_2\text{PO}_4^-$ ) and the diabolic ( $\text{HPO}_4^{2-}$ ) ions (Bhattacharyya and Jha, 2012). To overcome the P deficiency in soils, there are frequent applications of phosphatic fertilizers in agricultural fields. Plants absorb fewer amounts of applied phosphatic fertilizers and the rest is rapidly converted into insoluble complexes in the soil (Mckenzie and Roberts, 1990). Likewise, phosphate solubilizing bacteria have the capability to solubilize the residual or fixed soil P, increase the availability of P in the soil produce growth promoting substances (Selvakumar *et al.*, 2009), and thereby increase the overall P- use efficiency of the crops. Phosphate solubilizing microorganism can work efficiently to enhance the fraction of available phosphate to plants by solubilizing inorganic as well as organic phosphates (Saxena and Sharma, 2007). This process of phosphates solubilization is



associated to the production and release of organic acids of low molecular weight. It is accepted fact that their carboxyl and hydroxyl groups inter-chelate in the phosphate bound cations, resulting in their conversion to soluble forms (Alam *et al.*, 2002). Phosphate solubilizing bacteria are also produce plant growth hormones, participate in bio control activities and effect the process of nitrogen fixation (Neelam and Meenu., 2003). *Bacillus*, *Achromobacter*, *Agrobacterium* *Micrococcus*, *Burkholderia*, *Erwinia*, *Flavobacterium*, *Pseudomonas*, and *Rhizobium*, are some well-known phosphate-solubilizing bacterial strains which are used as biofertilizers and bio-control agents for agriculture improvement (Tamilarasi *et al.*, 2008; Srivastava and Shalini 2009).

Sundara *et al.* (2002) applied rock phosphate with a PSB (*Bacillus megaterium* var. phosphaticum) in lignite-based culture medium in a field experiment. They found that without P application PSB amendment could increase sugarcane yield by 12.6 percent. PSB and P fertilizer together reduced the P requirement by 25 percent. PSB also improved the sugar yield and juice quality. In conclusion, PSB may be of greatest value in allowing use of cheaper P sources (e.g., rock phosphate instead of superphosphate). Singh and Pareek (2003) reported that combined inoculation of *Rhizobium* + PSB significantly increased the nitrogen and phosphorus content in grain and stover, N and P uptake kg ha<sup>-1</sup> over control. Yadav *et al.* (2007) observed that highest grain yield (12.49 q ha<sup>-1</sup>) grains pod<sup>-1</sup> (13), test weight (42 g) and maximum number of nodules (36) were recorded with application of *Rhizobium* and PSB + P<sub>2</sub>O<sub>5</sub> @ 75 kg ha<sup>-1</sup> + poultry manure at 5 t ha<sup>-1</sup> in green gram. Singh *et al.* (2004) conducted a field experiment on sandy loam soil at Jobner and revealed that the both *Rhizobium* and PSB inoculation significantly increased the N, P content and its uptake in grain and Stover of greengram over single inoculation.

## **Mycorrhizae**

Symbiosis between plant roots and certain soil fungi e.g. Vesicular Arbuscular Mycorrhiza (VAM) plays an important role in phosphorus cycling and its uptake by plants (Biswas *et al.*, 2001). These symbiotic micro-organisms have extensive mycelial network and can increase the transport of other mineral elements such as zinc and copper. VAM fungi can play an important role in enhancing P availability to plants in deficient soils and can save P-fertilizer by 25-30% (Somani *et al.*, 1990). Additional benefits from the mycorrhizal symbiosis include increased tolerance of heavy metal contamination or drought, as well as lesser

---

susceptibility to root pathogens or herbivory. Mycorrhizal fungi may also improve soil quality by having a direct influence on soil aggregation (Rillig et al., 2002) and therefore aeration and water dynamics. This is of two types, the ectomycorrhizae, is found in trees and is beneficial for forest trees whereas, the second one is endomycorrhizae which is common in crop plants.

In India, Tata Energy Research Institute (TERI), New Delhi and Forest Research Institute, Dehradun have established mycorrhizae banks. Inocula of these can be procured as needed and used in horticulture and forestry programmes.

### **Silicate solubilizing bacteria (SSB)**

Microorganisms are capable of degrading silicates and aluminum silicates. During the metabolism of microbes several organic acids are produced and these have a dual role in silicate weathering. The studies conducted with a *Bacillus* sp. isolated from the soil of granite crusher yard showed that the bacterium is capable of dissolving several silicate minerals under *in vitro* condition. The examination of anthropogenic materials like cement, agro inputs like super phosphate and rock phosphate exhibited silicate solubilizing bacteria to a varying degree. The bacterial isolates made from different locations had varying degree of silicate solubilizing potential. Rice responded well to application of organic siliceous residue like rice straw, rice husk and black ash @ 5 t/ha. Combining SSB with these residues further resulted in increased plant growth and grain yield. This enhancement is due to increased dissolution of silica and nutrients from the soil.

### **Zinc solubilizers**

Zinc being utmost important is found in the earth's crust to the tune of 0.008 per cent but more than 50 per cent of Indian soils exhibit deficiency of zinc with content must below the critical level of 1.5 ppm of available zinc (Katyal and Rattan, 1993). The plant constraints in absorbing zinc from the soil are overcome by external application of soluble zinc sulphate ( $ZnSO_4$ ). But the fate of applied zinc in the submerged soil conditions is pathetic and only 1-4% of total available zinc is utilized by the crop and 75% of applied zinc is transformed into different mineral fractions (Zn-fixation) which are not available for plant absorption (crystalline iron oxide bound and residual zinc) (Alloway, 2008). The zinc can be solubilized by

microorganisms viz., *B. subtilis*, *Thiobacillus thiooxidans* and *Saccharomyces* sp. These microorganisms can be used as bio-fertilizers for solubilization of fixed micronutrients like zinc (Raj, 2007). The results have shown that a *Bacillus* sp. (Zn solubilizing bacteria) can be used as bio-fertilizer for zinc or in soils where native zinc is higher or in conjunction with insoluble cheaper zinc compounds like zinc oxide (ZnO), zinc carbonate (ZnCO<sub>3</sub>) and zinc sulphide (ZnS) instead of costly zinc sulphate (Mahdi et al. 2010).

### **Plant Growth Promoting Rhizobacteria (PGPR)**

The group of bacteria that colonize roots or rhizosphere soil and beneficial to crops are referred to as plant growth promoting rhizobacteria (PGPR) (Subbarao 1999; Wu *et al.*, 2005; Heidari *et al.*, 2011). Alternatively, Somers et al. (2004) classified PGPR based on their functional activities as (i) biofertilizers (increasing the availability of nutrients to plant), (ii) phytostimulators (plant growth promotion, generally through phytohormones), (iii) rhizoremediators (degrading organic pollutants) and (iv) biopesticides (controlling diseases, mainly by the production of antibiotics and antifungal metabolites) (Antoun and Pre´vost, 2005). Furthermore, in most studied cases, a single PGPR will often reveal multiple modes of action including biological control (Kloepper, 2003; Vessey, 2003). However, in accordance with their degree of association with the plant root cells, PGPRs can be classified into extracellular plant growth promoting rhizobacteria (ePGPR) and intracellular plant growth promoting rhizobacteria (iPGPR). The ePGPRs may exist in the rhizosphere, on the rhizoplane or in the spaces between the cells of root cortex. For eg. *Agrobacterium*, *Arthrobacter*, *Azotobacter*, *Azospirillum*, *Bacillus*, *Burkholderia*, *Caulobacter*, *Chromobacterium*, *Erwinia*, *Flavobacterium*, *Micrococcous*, *Pseudomonas* and *Serratia* etc. iPGPRs locates generally inside the specialized nodular structures of root cells. For eg. *Allorhizobium*, *Azorhizobium*, *Bradyrhizobium*, *Mesorhizobium* and *Rhizobium* of the family *Rhizobiaceae*. The potentiality of PGPR in agriculture is steadily increased as it offers an attractive way to replace the use of chemical fertilizers, pesticides and other supplements. Several PGPR formulations are currently available as commercial products for agricultural production of beneficial crops. Bertrand et al. (2000) showed that a rhizobacterium belonging to the genus *Achromobacter* could enhance root hair number and length in oilseed rape (*Brassica napus*). *Achromobacter* increased NO<sub>3</sub> and K uptake and, consequently, shoot and root dry weights by 22 to 33 percent and 6 to 21 percent respectively.

---

## **K-Solubilizing Bacteria**

Bacteria such as *Fratureia aurantia* are capable of mobilizing mixture of K into a usable form to the plants known as K solubilizing bacteria, applied to all crops in association with other Biofertilizers without any antagonistic effect. However, while positive responses have been observed in a wide range of field trials, there is remarkable inconsistency in responses across crops, regions and other conditions (Ghosh, 2004).

## **Conclusion**

With excessive use of chemical fertilizer, flora and fauna is adversely effected as well as it disturb the ecological balance. Indian soil fertility is diminishing gradually. In some way biofertilizer can help to provide nutritional requirement of agricultural crop. The major challenge in this area of research lies in the fact that identification of local and efficient strain of particular area that can survive better in competition with other microorganism. The integration of different microbial capabilities into combined biofertilizers with numerous potential yield-promoting effects is also desirable in integrated nutrient management.

## **REFERENCE:**

- Ahemad, M. and Kibret, M. (2014). *Mechanisms and applications of plant growth promoting rhizobacteria: Current perspective Journal of King Saud University-Science, Vol 26, pp.1-20.*
- Alam, S., S. Khalil, N. Ayub, and M. Rashid. (2002). *In vitro Solubilization of Inorganic Phosphate by Phosphate Solubilizing Microorganisms (PSM) from Maize Rhizosphere. J. Agri. Bio., Vol 4 pp. 454–458.*
- Alloway, B. J. (2008). *Zinc in soils and crop nutrition. Second edition, IZA and IFA publishers, Brussels, Belgium and Paris, France. Pp. 21-22.*
- Antoun, H. and Pre´vost, D. (2005). *Ecology of plant growth promoting rhizobacteria. In: Siddiqui, Z.A. (Ed.), PGPR: biocontrol and biofertilization, Springer, Dordrecht, pp.1–38.*

- Araujo, A. S. F., Santos, V.B. and Monteiro, R.T.R. (2008). Responses of soil microbial biomass and activity for practices of organic and conventional farming systems in Piauí state, Brazil. *Eur J Soil Biol.*, Vol 44 pp. 225–230.
- Bahadur, L., Tiwari, D.D., Mishra, J. and Gupta, B.R. (2013). Nutrient management in rice-wheat sequence under sodic soil. *Journal of the Indian Society of Soil Science*, Vol 61, pp. 341-346.
- Balachandran, D. and Nagarajan, P. (2002). Dual inoculation of *Rhizobium* and phosphobacteria with phosphorus on blackgram cv. Vamban 1. *Madras Agric. J.* Vol 89 No.10- 12, pp. 691-693.
- Bertrand, H., Plassard, C., Pinochet, X., Touraine, B., Normand, P. and J.C. Cleyet-Marel, (2000). Stimulation of the ionic transport system in *Brassica napus* by a plant growth promoting rhizobacterium (*Achromobacter* sp.). *Can J Microbiol.*, Vol 46, pp. 229–236.
- Bhardwaj, D., Ansari, M.W., Sahoo, R.K. and Tuteja, N. (2014). Biofertilizers function as key player in sustainable agriculture by improving soil fertility, plant tolerance and crop productivity. *Microbial Cell Factories*, Vol 13, pp.66.
- Bhattacharyya, P. N. and Jha, D. K. (2012). Plant growth-promoting rhizobacteria (PGPR):
- Biswas, B.C. Yadav, D.S., and Maheshwari, S. (1985). Bio-fertilizers in Indian Agriculture. *Fertilizer News*, Vol 30 No. 10, pp. 20-28.
- Biswas, B.C., Das, S. and Subhash, K.P. (2001). Crop response to bio-fertilizers. *Fertilizers News*, Vol 46, pp. 15-24.
- emergence in agriculture. *World J Microbiol Biotechnol*, Vol 28 pp.1327–1350.
- Ezawa, T. Smith, S. E. and Smith, F.A. (2002). P Metabolism and Transport in AM Fungi. *Plant and Soil*, Vol 244 No.1-2, pp. 221-230.
- Gaur, A.C. (1991). Phosphate solubilizing micro-organism and bio-fertilizers. Omega Scientific Publishers, New Delhi, pp. 176.
- Ghosh, N. (2004). Promoting Biofertilisers in Indian Agriculture. *Economic and Political Weekly*, Vol 5, pp. 5617-5625.
- Heidari, M., Mousavinik, S. M. and Golpayegani, A. (2011). Plant Growth Promoting Rhizobacteria (PGPR) Effect on Physiological Parameters and Mineral Uptake in Basil
-

- (*Ocimum basilicum* L.) Under Water Stress. *ARPJ Journal of Agricultural and Biological Science*, Vol 6 No.5, pp. 6-11.
- Jain, A., Kumar, S. and Panwar, J. D. S. (2007). Response of mungbean to phosphorus and micronutrient on N and P uptake and seed quality. *Legume Res.* , Vol 30 No. 3, pp. 201-204.
- Kannaiyan, S. (2002). *Biofertilizers for sustainable crop production. Biotechnology of Biofertilizers.* Narosa Publishing House, New Delhi, India, pp. 9-49.
- Katyal, J. C. and Rattan, R. K. (1993). Distribution of zinc in Indian soils. *Fert. News*, Vol 38 No3, pp. 15-26.
- Katyal, J. C., Venkateshwarlu, B. and Das, S. K. (1994). Biofertilizer for Nutrient Supplementation in Dryland Agriculture. *Fertiliser News*, Vol 39 No 4 pp. 27-32.
- Khan, M. S., Zaidi, A., Wani, P. A. and Oves, M. (2009). Role of plant growth promoting rhizobacteria in the remediation of metal contaminated soils. *Environ. Chem. Lett.*, Vol 7, pp. 1–19.
- Khan, T. A., Amani, S. and Naeem, A. (2012a). Glycation promotes the formation of genotoxic aggregates in glucose oxidase. *Amino Acids*, Vol 43 No 3, pp. 1311-1322.
- Kloepper, J. W. (2003). A review of mechanisms for plant growth promotion by PGPR In: Reddy, M.S., Anandaraj, M., Eapen, S.J., Sarma, Y.R., Kloepper, J.W. (Eds.), *Abstracts and Short Papers.*
- Kucey, R.M.N., Janzen, H.H. and Legett, M.E. (1989). Microbially mediated increases in plant available phosphorus. *Adv. Agron.*, Vol 42, pp. 199–228.
- Kumar, V. and Ahlawat, I.P. S. (2004). Carry over effect of biofertilizers and nitrogen applied to wheat (*Triticum aestivum*) and direct applied N in maize (*Zea mays*) in wheat-maize cropping system. *Indian Journal of Agronomy*, Vol 49, pp. 233-236.
- Lugtenberg, B. J. J., Chin-A-Woeng, T. F. C. and Bloemberg, G. V. (2002). "Microbe-plant interactions: principles and mechanisms," *Antonie Van Leeuwenhoek*, Vol. 81No. 1–4, pp. 373–383.
- Mahdi, S.S., Dar, S. A., Ahmad S. and Hassan, G. I. (2010). Zinc availability- A major issue in agriculture. *Research Journal Agricultural Sciences*, Vol 3 No.3 pp. 78-79.
- Mahdi, S.S., Hassan, G.I., Samoon, S.A., Rather, H.A., Dar, S.A. and Zehra, B. (2010). Biofertilizers in organic agriculture. *J. Phytol.*, Vol 2 No.10 pp.42-54.

- Mazid, M., Khan, T. A. and Mohammad, F. (2011d). Response of crop plants under sulphur stress tolerance: A holistic approach. *Journal of Stress Physiology and Biochemistry*, Vol 7 No. 3 pp. 23-57.
- Mazid, M., Khan, T. A. and Mohammad, F. (2011f). Cytokinins, A classical multifaceted hormone in plant system. *Journal of Stress Physiology & Biochemistry*, Vol 7 No. 4 pp. 347-368.
- Mazid, M., Khan, T. A. and Mohammad, F. (2012b). Role of NO in H<sub>2</sub>O<sub>2</sub> regulating responses against temperature and ultraviolet induced oxidative stress in plants. *Acta Biologica Indica*, Vol 1 No.1, pp. 1-16.
- Mohammadi, K. and Yousef, S. (2012). Bacterial Biofertilizers for sustainable crop Production: A Review. *ARPN Journal of Agricultural and Biological Science*, Vol. 7 No.5, pp.307-316.
- Naeem, A., Khan, T. A., Muzaffar, M., Ahmad, S. and Saleemuddin, M. (2011). A partially folded state of ovalbumin at low pH tends to aggregate. *Cell Biochemistry and Biophysics*, Vol 59 pp. 29-38.
- Neelam, T. and Meenu, S. (2003). Phosphate solubilization, exopolysaccharide production and indole acetic acid secretion by rhizobacteria. *Indian J. Microbiol.*, Vol 43, pp. 37-40.
- Nina, K., Thomas, W. K. and Prem, S. B. (2014). Beneficial organisms for nutrient uptake. VFRC report 2014/1, virtual fertilizer research center. Washington, DC: Wageningen Academic Publishers; pp. 63.
- Patil, P., Ghag, P. and Patil, S. (2013). Use of Bio-fertilizers and Organic Inputs - as LISA technology by farmers of Sangamner. *International Journal of Advancements in Research & Technology*, Vol 2, No. 7.
- Rahman, M. M., Amano, T. and Shiraiwa, T. (2009). Nitrogen use efficiency and recovery from N fertilizer under rice-based cropping systems. *Australian Journal of Crop Science Southern Cross Journals*, Vol 3 No 6, pp. 336-351.
- Raj. S. A. (2007). Bio-fertilizers for micronutrients. *Biofertilizer Newsletter* (July), pp 8-10.
- Rana, A., Joshi, M., Prasanna, R., Shivay, Y. S. and Nain, L. (2012). Biofortification of wheat through inoculation of plant growth promoting rhizobacteria and cyanobacteria. *European Journal of Soil Biology* Vol 50, pp. 118-26.

- Rillig, M. C., Wright, S. F. and Eviner, V. (2002). *The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: comparing effects of five plant species. Plant Soil, Vol 238 pp. 325-333.*
- Sahoo, R.K., Ansari, M.W., Dangar, T.K., Mohanty, S. and Tuteja, N. (2013). *Phenotypic and molecular characterization of efficient nitrogen fixing Azotobacter strains of the rice fields. Protoplasma, doi:10.1007/s00709-013-0547-2.*
- Saikia, S .P. and Jain, V. (2007). *Biological nitrogen fixation with non-legumes: An achievable target or a dogma. Current Science, Vol 92 No. 3, pp. 317- 322.*
- Saxena, A. and Sharma, J. V. (2007). *Isolation of tricalcium phosphate solubilizing strains from semiarid agricultural fields of Rajasthan, India. J. Pure Appl. Microbiol., Vol 1 pp. 269-280.*
- Selvakumar, G., Mohan, M., Kundu, S., Gupta, A. D., Joshi, P., Nazim, S. and Gupta, H. S. (2008). *Cold tolerance and plant growth promotion potential of Serratia marcescens strain SRM (MTCC 8708) isolated from flowers of summer squash (Cucurbita pepo). Lett. Appl. Microbiol., Vol 46, pp. 171–175.*
- Sepat, R. K., Rai, R. K. and Dhar, S. (2010). *Planting systems and integrated management for enhanced wheat (Triticum aestivum) productivity. Indian Journal of Agronomy, Vol 55, pp. 114-118.*
- Shrivastava, R. and Shalini (2009). *Antifungal activity of Pseudomonas fluorescens against pathogenic fungi. The internet j. Microbiol., Vol 7 No. 2.*
- Singh, A. P., Tripathi, M.K. and Singh, S. (2004). *Growth and yield of greengram as influenced by biofertilizer and phosphorus application. Ann. Biol., Vol 20 No. 2, pp. 227-232.*
- Singh, B. and Pareek, R.G. (2003). *Studies on phosphorus and bio- inoculants on biological nitrogen fixation, concentration, uptake, quality and productivity of mungbean. Ann. Agric. New series., Vol 24 No.3, pp. 537-541.*
- Singh, G., Choudhary, P., Meena, B. L., Rawat, R. S. and Jat, B. L. (2016). *Integrated nutrient management in blackgram under rainfed condition. International Journal of Recent Scientific Research, Vol 7 No. 10, pp. 13894-13895.*
- Singh, J.S., Pandey, V.C. and Singh, D.P. (2011). *Efficient soil microorganisms: a new dimension for sustainable agriculture and environmental development. Agric Ecosyst Environ, Vol 140, pp. 339–353.*
-



- Sinha, R.K., Valani, D., Chauhan, K. and Agarwal, S. (2014). *Embarking on a second green revolution for sustainable agriculture by vermiculture biotechnology using earthworms: reviving the dreams of Sir Charles Darwin*. *Int J Agric Health Saf* , Vol 1, pp. 50–64.
- Somani, L. L., Bhandari, S. C., Saxena, S. N. and Vyas, K. K. (Ed.) (1990). *Bio-fertilizers*. Scientific Publication, Jodhpur Pp. 142-145.
- Somers, E., Vanderleyden, J. and Srinivasan, M,( 2004). *Rhizosphere bacterial signalling: a love parade beneath our feet*. *Crit. Rev. Microbiol.*, Vol 30, pp 205–240.
- Subba Rao, N. S. (1999). *Soil Microbiology (Fourth Edition of Soil Microorganisms and Plant Growth)*. Science Publishers, Inc. USA.
- Swarup, A., Damodar, Reddy, D. and Prasad, R. N. (Eds.) (1998). *Proceedings of National Workshop on Longterm Soil Fertility Management through Integrated Plant Nutrient Supply*. Indian Institute of Soil Science, Bhopal, pp. 335.
- Tamilarasi S., Lakshmanaperumalsamy, P., Nanthakumar, K. and Karthikeyan, K. (2006). *Diversity of root associated microorganisms of selected medicinal plants and influence of rhizo microorganisms on the antimicrobial property of Coriandrum Sativum J*. *Environmental Biology*, Vol. 29 No. 1 pp. 127-134.
- Tamilnadu Agricultural University, (2009). Department of agricultural meteorology. (<http://www.tnau.ac.in/sems/Agmet/weather.html> accessed on 10.12.2009)
- Vance, C. P. (1997). *Biological fixation of N<sub>2</sub> for ecology and sustainable agriculture*. Springer-Verlag, pp. 179.
- Venkatashwarlu, B. (2008). *Role of bio-fertilizers in organic farming: Organic farming in rain fed agriculture: Central institute for dry land agriculture, Hyderabad*, pp. 85-95.
- Verma, L. N. (1993). *Biofertiliser in agriculture*. In: *Organics in soil health and crop production* (Thampan, P.K., ed.). Peekay Tree Crops Development Foundation, Cochin, India. pp. 152-183.
- Vessey, J. K. (2003). "Plant growth promoting rhizobacteria as biofertilizers," *Plant and Soil*, Vol. 255, No. 2, pp. 571–586.

- Wani, S. P. and Lee, K. K. (1995). *Microorganisms as biological inputs for sustainable agriculture in Organic Agriculture (Thampan, P.K.ed.) Peekay Tree Crops Development Foundation, Cochin, India. Pp-39-76.*
- Wood, T. and Cummings, B. (1992). *Biotechnology and the future of VAM commercialization. In: Mycorrhizae in Sustainable Agriculture. (ed GJ Bethlenfalvay, RG Linderman.). American Society of Agronomy Special Publication, Vol 54 pp. 468–487.*
- Wu, S. C., Cao., Z. H., Li., Z .G., Cheung, K. C. and Wong, M. H. (2005). *Effects of biofertilizer containing N-fixer, P and K solubilizers and AM fungi on maize growth: a greenhouse trial. Geoderma. Vol 125 pp. 155–166.*
- Yadav, A. K., Varghes, K. and Abraham, T. (2007). *Response of biofertilizers poultry manure and different levels of phosphorus on nodulation and yield of greengram (Vigna radiata L.) Cv. K-851. Agric. Sci. Digest., Vol 27 No. 3 pp. 212-215.*
- Youssef, M. M. A. and Eissa, M. F. M. (2014). *Biofertilizers and their role in management of plant parasitic nematodes. A review. E3 J Biotechnol. Pharm Res, Vol 5 pp.1–6.*