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## ROLE OF OXIDATIVE STRESS IN VITAMIN-D DEFICIENCY INDUCED MUSCLE WASTING

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### ABSTRACT

The effects and mechanisms of vitamin D activities in human skeletal muscle, as well as the existence of the vitamin D receptor in mature human skeletal muscle, are yet unknown. The major goal of this research is to create a vitamin D deficient rat model and investigate total protein breakdown in that strain. A variety of muscle-wasting illnesses have been linked to oxidative stress. The ordinarily well-balanced management of oxidant production and antioxidant activity is disrupted in oxidative stress. Enzymatic or chemical processes that create superoxide anion, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), or nitric oxide (NO) are sources of oxidants (NO). This study also looks at the role of distinct proteolytic processes in vitamin D deficiency-induced muscle atrophy. The study will have shown, to the best of our knowledge, that vitamin D deficiency causes muscle protein breakdown through upregulating the ubiquitin protein pathway. Increased muscle protein breakdown will result in a loss of muscle mass, which will be linked to a decrease in myogenic genes and an increase in atrophy marker genes.

**KEYWORDS:** Oxidative Stress, Vitamin-D Deficiency, Muscle Wasting, human skeletal muscle

### INTRODUCTION

Vitamin D in its hormonally dynamic structure, 1',25-dihydroxyvitamin D [1',25(OH)<sub>2</sub>D; calcitriol] is not only a controller of calcium and phosphate homeostasis, but it also has a variety of extra skeletal impacts, as shown by a variety of research findings from the past few years. These findings have been shown by a variety of researchers. These include the significant impact

that the vitamin D hormone has on the cardiovascular system, the focused sensory system, the endocrine system, and the immunological system, as well as the impact that it has on cell division and cell proliferation. The term "vitamin D" actually refers to two different compounds, both of which play an important role in the human body. Vitamin D<sub>2</sub> (ergocalciferol), which is produced in some plants but primarily in organisms, and vitamin D<sub>3</sub> (cholecalciferol), which is produced by human skin when exposed to daylight through the action of ultraviolet light on 7-dehydrocholesterol, are both included in this category. This photo-production is influenced by a few different circumstances, including age, UV exposure (scope, season, utilisation of sunscreens, and apparel), nationality (skin pigmentation), and scope of exposure. Each of these conditions has the potential to hinder the body's ability to synthesise the bioavailable form of vitamin D. Insufficient production of vitamin D<sub>3</sub> is also linked to a lack of time spent outside and poor behaviours in general, both of which contribute to an unhealthy lifestyle. Alternately, pre-vitamin D and vitamin D<sub>3</sub> may be converted into inert photoproducts if they are exposed to an excessive amount of sunshine. The formation of the bioactive form of vitamin D requires participation from a variety of organs. In order to begin, vitamin D<sub>3</sub> must first go through two stages of hydroxylation in the liver and the kidney. The ultimate product, hormonally active 1,25-dihydroxyvitamin D<sub>3</sub> (calcitriol), is produced as a consequence of the flow to its target tissues and works in a manner that is either genomic or non-genomic.

## **VITAMIN-D**

One of the most important roles of vitamin D is to regulate calcium and phosphate balance and bone development and maintenance. The 1,25-dihydroxyvitamin D [1,25(OH)<sub>2</sub>D<sub>3</sub>] hormone's physiologically dynamic structure binds to the vitamin D receptor (VDR), a member of the atomic receptor superfamily (2, 3). All over again, protein synthesis is the outcome of 1, 25(OH)<sub>2</sub>D<sub>3</sub> binding to its receptor (4). It has long been accepted that the gut, kidney, bone, and parathyroid glands are the primary targets of vitamin D supplementation (1). Vitamin D has been shown to have a key role in a wide range of tissues in the body, including skeletal muscle, during the last several decades (2). Vitamin D insufficiency causes muscular weakness in osteomalacia in adults and hypotonia in infants, according to clinical observations of the link between vitamin D and muscle function (5, 6). Myopathy linked to severe vitamin D insufficiency has been shown (7). The administration of modest doses of vitamin D to elderly participants decreased the number of hip fractures and falls (8). Using muscle biopsy samples taken from adults with

vitamin D insufficiency and proximal myopathy, he has discovered a power of type II muscle fibre degradation in eight out of eleven cases. The identification of the VDR in muscle cells provided more evidence of vitamin D's direct role in muscle tissue.

Non-genomic processes, lasting from a few seconds to a few minutes, may also be activated by Vitamin D3. Activation of adenylyl cyclase-cAMP-protein kinase signal transduction pathways is included in this system. An and phospholipase C-diacylglycerol-inositol (1,4,5)-trisphosphate-protein kinase C signal transduction pathways are involved. Since they may be involved in cross-talk with the core, the second couriers Raf (rapidly accelerated fibrous sarcoma)/MAPK are of particular importance. Among the earliest and most notable nongenomic activations is "transcaltachia," which is the rapid acceleration of intestinal calcium transport. The chondrocytes of the bone growth plate and the keratinocytes of the skin were shown to be affected by this effect. A different design of the VDR was used to identify agonists suitable to drive nongenomic consequences during the identification of the vitamin D3 receptor. Vitamin D3 and MARRS (a film-related rapid reaction steroid binding protein) interaction has also been studied in detail in this study. The receptors for kinases, phosphatases, and ion channels are located in the layer within caveolae/lipid pontoons.

The immune system is strongly modulated by vitamin D3 (cholecalciferol). For example, keratinocytes that form the mucocutaneous barrier increase VDR and 1-hydroxylase expression following a skin injury to enhance immune responses. After a Mycobacterium TB infection or exposure to lipopolysaccharides, monocytes and macrophages behave similarly. In all circumstances, increased synthesis of cathelicidin and -defensin 2 might demonstrate antimicrobial effects, enhancing natural resistance. Self-created vitamin D3 may be delivered by monocytes or macrophages to operate locally on started T and B cells, which can independently regulate cytokine and immunoglobulin production. The many effects of vitamin D3 on the immune system play a critical role in the fight against incurable illnesses. Tuberculosis and other viral diseases of the upper respiratory tract are especially vulnerable to the natural resistance that vitamin D3 confers. Vitamin D has been shown to have antibacterial properties, and its shortage has been shown to have a negative impact on overall well-being and lifespan. An infection risk may be reduced in numerous ways, including balancing antimicrobial production, controlling local immunological and immune-inflammatory reactions, increasing the lethality of attacking live organisms, and directing local immune and immune-inflammatory responses. As a result,

vitamin D provides a small preventive and maybe a restorative alternative, either alone or in conjunction with standard medications. The frequency of autoimmune disorders such as type 1 diabetes, multiple sclerosis, and Crohn's disease has been linked to sunshine exposure and subsequent vitamin D<sub>3</sub> synthesis, according to biological studies. Inflammatory bowel disease (IBD), rheumatoid arthritis (RA), and type 1 diabetes have all shown a rise in prevalence at greater levels. Corticosteroid usage in children with asthma seems to have a direct correlation with vitamin D levels in the bloodstream.

### **1 Vitamin D: Production, Metabolism And Mechanisms Of Action**

“Vitamin D comes in two forms (D<sub>2</sub> and D<sub>3</sub>) which differ chemically in their side chains. These structural differences alter their binding to the carrier protein vitamin D binding protein (DBP) and their metabolism, but in general the biologic activity of their active metabolites is comparable. Vitamin D<sub>3</sub> is produced in the skin from 7-dehydrocholesterol by UV irradiation, which breaks the B ring to form pre-D<sub>3</sub>. Pre-D<sub>3</sub> isomerizes to D<sub>3</sub> but with continued UV irradiation to tachysterol and lumisterol. D<sub>3</sub> is preferentially removed from the skin, bound to DBP. The liver and other tissues metabolize vitamin D, whether from the skin or oral ingestion, to 25OHD, the principal circulating form of vitamin D. Several enzymes have 25-hydroxylase activity, but CYP2R1 is the most important. 25OHD is then further metabolized to 1,25(OH)<sub>2</sub>D principally in the kidney, by the enzyme CYP27B1, although other tissues including various epithelial cells, cells of the immune system, and the parathyroid gland contain this enzyme. 1,25(OH)<sub>2</sub>D is the principal hormonal form of vitamin D, responsible for most of its biologic actions. The production of 1,25(OH)<sub>2</sub>D in the kidney is tightly controlled, being stimulated by parathyroid hormone (PTH), and inhibited by calcium, phosphate and FGF23. Extrarenal production of 1,25(OH)<sub>2</sub>D as in keratinocytes and macrophages is under different control, being stimulated primarily by cytokines such as tumor necrosis factor alfa (TNF $\alpha$ ) and interferon gamma (IFN $\gamma$ ). 1,25(OH)<sub>2</sub>D reduces 1,25(OH)<sub>2</sub>D levels in cells primarily by stimulating its catabolism through the induction of CYP24A1, the 24-hydroxylase. 25OHD and 1,25(OH)<sub>2</sub>D are hydroxylated in the 24 positions by this enzyme to form 24,25(OH)<sub>2</sub>D and 1,24,25(OH)<sub>3</sub>D, respectively. This 24-hydroxylation is generally the first step in the catabolism of these active metabolites to the final end product of calcitroic acid, although 24,25(OH)<sub>2</sub>D and 1,24,25(OH)<sub>3</sub>D has their own biologic activities. CYP24A1 also has 23-hydroxylase activity that leads to a different end product. Different species differ in their ratio of 23-hydroxylase/24-hydroxylase

activity in their CYP24A1 enzyme, but in humans the 24-hydroxylase activity predominates. Like CYP27B1, CYP24A1 is widely expressed. CYP24A1 is induced by 1,25(OH)<sub>2</sub>D in most tissues, which serves as an important feedback mechanism to avoid vitamin D toxicity. In the kidney, PTH inhibits CYP24A1, whereas FGF23, calcium and phosphate stimulate it, just the opposite of the actions of these hormones and minerals on CYP27B1. However, such regulation is not seen in other tissues. In macrophages, CYP24A1 is either missing or defective, so in situations such as granulomatous diseases like sarcoidosis in which macrophage production of 1,25(OH)<sub>2</sub>D is increased, hypercalcemia and hypercalciuria due to elevated 1,25(OH)<sub>2</sub>D can occur without the counter regulation by CYP24A1.”

### **Metabolism**

In order for vitamin D<sub>3</sub> to be effective, it must be further digested once it is created in the epidermis. The first step, 25-hydroxylation, is predominantly carried out in the liver, although this enzyme activity may also be found in other organs. As we'll see in a moment, the 25-hydroxylase family is rather large. Vitamin D is most often found in the form of 25OHD. 1,25(OH)<sub>2</sub>D is the most powerful metabolite of vitamin D and accounts for most of its biological activities. However, in order for vitamin D metabolites to acquire optimum biological activity, they must be further hydroxylated in the 1 position by the enzyme CYP27B1. While the 25-hydroxylase is found in various tissues, the 1 hydroxylation occurs exclusively in the kidney. It is possible for 25OHD and 1,25(OH)<sub>2</sub>D to be hydroxylated at all 24 places of the molecule. Because 1,25(OH)<sub>2</sub>D and 1,24(OH)<sub>2</sub>D have comparable biologic potency and 1,24,25(OH)<sub>3</sub>D has activity around 1/10 that of 1,25(OH)<sub>2</sub>D, this might activate the metabolite or analogue. Metabolites having an existing 25OH group, on the other hand, undergo 24-hydroxylation, resulting in additional degradation. The specifics of these responses are outlined in the next paragraphs.

### **2 Cutaneous Production of Vitamin D<sub>3</sub>**

Among the Kandutsch-Russell cholesterol pathways is the vitamin D precursor 7-dehydrocholesterol (DHC). 7-dehydrocholesterol reductase, the enzyme that converts 7-DHC to cholesterol, is controlled by a variety of variables, including vitamin D and cholesterol that facilitate the breakdown of 7-DHC therefore allowing for larger quantities of 7-DHC to be converted to vitamin. However, while 7-DHC irradiation produced pre-D<sub>3</sub> (which later undergoes thermal rearrangement of the triene structure to generate D<sub>3</sub>), lumisterol and

tachysterol, the physiologic control of this pathway was not well understood until the work of Holick and his collaborators. In the presence of UV or solar irradiation (with maximum effective wavelengths between 290 and 310), they found that pre-D3 is rapidly formed and may reach its maximum concentration within hours. Lumisterol and Tachysterol are formed by UV irradiation of pre-D3. Both the degree of pigmentation on the epidermis and the intensity of the exposure are linked to the amount of time needed to reach this maximum concentration of pre-D3, yet they have no effect on it. Pre-D3 levels peak, but the lumisterol, which is not physiologically active, continues to build over time. In addition to pre-D3, tachysterol is also produced, although it does not build with prolonged exposure to UV radiation. As pre-D3 levels decline, lumisterol may be turned back to pre-D3 by the body. In the absence of D3, pre-D3 is progressively transformed to D3 at 37°C. Because of the delayed thermal conversion of pre-D3 to D3 and the conversion of lumisterol to pre-D3, brief exposure to sunlight is predicted to result in a prolonged synthesis of D3 in the exposed skin. Toxic D3 would not be produced by prolonged exposure to sunlight because of the photoconversion of pre-D3 to lumisterol and the photoconversion of D3 itself to suprasterols I and II and 5,6transvitamin D3.

Melanin in the epidermis may inhibit sunlight's ability to produce D3 in the skin by absorbing UV irradiation. Because of this, people of African and Hispanic descent who live in temperate climates may have lower levels of 25OHD (a well-documented proxy for vitamin D levels in the body). As a result of sunlight exposure, melanin formation rises, and therefore another strategy for preventing excess D3 production is available. The intensity of UV irradiation is also critical to the synthesis of D3. With greater 25OHD concentrations during the summer and lower levels during the winter, there may be significant seasonal change. This seasonal variation's magnitude is determined by the latitude and, as a result, by the amount of direct sunlight that is exposed to the skin. Edmonton, Canada (52°N) has a short window of time from mid-October to mid-April in which D3 production is minimal; Boston (42°N) has a somewhat longer window of time; while Los Angeles (34°N) and San Juan (18°N) have the ability to manufacture D3 all year long. Those results are applicable to the ocean floor. Skiers can synthesise vitamin D even on cloudy days in the winter because of the lower air absorption of UVB at higher altitudes. In the summer, the skin may produce more D3 than at any other time of year, with peak production occurring about midday. In locations where D3 generation is hindered by clothing and sunscreen, this is a good thing. That Bedouins, who wear full body gear, are more likely than Israeli Jews to suffer

from rickets and osteomalacia when exposed to the same amount of sunshine, is one possible reason.

### **3 Transport In Blood**

DBP (vitamin D binding protein) and albumin are the primary carriers of vitamin D metabolites in the blood (12-15 percent ). There is only around 2% saturation of DBP at typical values of 4-8 M, well beyond the level of vitamin D metabolites. Vitamin D metabolites have a strong affinity for DBP, which means that only 0.03 percent of 25OHD and 24,25(OH)2D and 0.4 percent of 1,25(OH)2D are free in normal conditions. The total 25OHD and 1,25(OH)2D levels will be lowered without necessarily impacting the free concentrations in conditions such liver disease and nephrotic syndrome that result in reduced DBP and albumin levels. Acute sickness also reduces DBP levels, which may make it difficult to evaluate total 25OHD levels. DBP levels were shown to be lower among African Americans in earlier investigations employing monoclonal antibodies, but these findings were not verified using polyvalent antibody-based tests. This may create hypercalcemia without necessarily elevating the overall quantities of 1,25(OH)2D, which can be caused by vitamin D intoxication.

To the vast majority of cells, it is impossible to access vitamin D metabolites that are coupled to DBP. Consequently, it is the unbound concentration, as proposed by the free hormone hypothesis, that is crucial for cellular absorption. Studies in mice with the DBP gene removed or in people with a mutation support the idea that DBP serves as a storage site for vitamin D metabolites, but that it is the free concentration that penetrates cells and exerts physiologic activity. The vitamin D metabolites in DBP knockout mice are likely entirely free or bioavailable. Vitamin D insufficiency is not evident in these mice until they are fed a vitamin D-deficient diet, despite the fact that their blood 25OHD and 1,25(OH)2D levels are very low. DBP knockout mice have normal 1,25(OH)2D levels and indicators of vitamin D action, such as intestinal TRPV6, calbindin 9k, PMCA1b, and renal TRPV5, in their tissues. A significant loss of the DBP gene's coding region (and the nearby NPPFR2 gene) has recently been discovered in a family. With vitamin D treatment, the proband had normal calcium, phosphate, and PTH levels, despite low 25OHD, 24,25(OH)2D, and 1,25(OH)2D levels (oral or parenteral). Free 25OHD levels were within acceptable limits. Between the proband and the normal sibling, the carrier sibling had vitamin D metabolite levels. DBP has been shown to act as a circulatory reservoir for vitamin D metabolites in both human and DBP null mouse investigations. There is

also a discussion over whether the free concentration of 25OHD, for example, is a better measure of vitamin D nutritional status than total 25OHD, because DBP levels, and hence total 25OHD levels, may be altered by liver illness, nephrotic syndrome, pregnancy, and inflammatory conditions. Vitamin D metabolites coupled to DBP may be transported into cells through the megalin/cubilin complex found in the kidney, placenta, and parathyroid glands. In addition to limiting renal losses, this may have a role in the delivery of vitamin D into the foetus as well as the control of PTH secretion. As it turns out, mice missing the megalin/cubilin complex have a shorter lifespan and show signs of osteomalacia, which suggests a function in vitamin D transport into cells involved in vitamin D signalling.

#### **4 Mechanism Of Action**

It is a transcription factor, the vitamin D receptor, that 1,25(OH)<sub>2</sub>D binds to (VDR). Genes that contain particular DNA sequences known as vitamin D response elements (VDREs) play an important role in the effects of 1,25(OH)<sub>2</sub>D. (VDREs). The gene has thousands of VDREs, some of which are hundreds of base pairs distant from the coding region of the gene. In contrast, some of the effects of 1,25(OH)<sub>2</sub>D are more rapid, and may be linked to a less well-known membrane-bound vitamin D receptor (VDR), as well as to the VDR operating outside of the nucleus. However, VDR's ligand 1,25(OH)<sub>2</sub>D is not required for all of its effects. In the last several years, we've learned a lot more about how VDR controls gene expression.

- **VDR and Transcriptional Regulation**

“The VDR was discovered in 1969 (although only as a binding protein for an as yet unknown vitamin D metabolite subsequently identified as 1,25(OH)<sub>2</sub>D), and was eventually cloned and sequenced in 1987. Inactivating mutations in the *VDR* result in hereditary vitamin D resistant rickets (HVDRR). Animal models in which the *VDR* has been knocked out have the full phenotype of severe vitamin D deficiency indicating that the VDR is the major mediator of vitamin D action. The one major difference is the alopecia seen in HVDRR and *VDR* knockout animals, a feature not associated with vitamin D deficiency, suggesting that the VDR may have functions independent of 1,25(OH)<sub>2</sub>D at least in hair follicle cycling. The VDR is a member of a large family of proteins (over 150 members) that includes the receptors for the steroid hormones, thyroid hormone, vitamin A family of metabolites (retinoids), and a variety of cholesterol metabolites, bile acids, isoprenoids, fatty acids and eicosanoids. A large number of family members have no known ligands, and are called orphan receptors. VDR is widely, although not



universally, distributed throughout the different tissues of the body. Many of these tissues were not originally considered target tissues for 1,25(OH)<sub>2</sub>D. The discovery of VDR in these tissues along with the demonstration that 1,25(OH)<sub>2</sub>D altered function of these tissues has markedly increased our appreciation of the protean effects of 1,25(OH)<sub>2</sub>D.”

- **Non-Genomic Actions**

The biological effects of many hormones that act as ligands for nuclear hormone receptors may be mediated via membrane receptors rather than their nuclear hormone receptors' corresponding receptors. Corticosteroids and thyroid hormone are examples of these hormones. Some studies have indicated that 1,25(OH)<sub>2</sub>D has a fast impact, independent of gene regulation, on a subset of cells. This action seems to be mediated by a distinct membrane receptor. Figure depicts a model for such effects. Calcium and chloride channel function, the activation of protein kinase C and distribution of phospholipase C have all been found to be controlled by 1,25(OH)<sub>2</sub>D in a variety of cell types including osteoblasts, muscle, and the digestive tract. In the gut, 1,25(OH)<sub>2</sub>D's fast effects have been most widely examined. To characterise the quick commencement of calcium flow through the gut of a vitamin D replete chick, Norman's team created the term transcaltachia. Actinomycin D pretreatment was unable to stop this enhanced flow, whereas voltage gated L type channel inhibitors and protein kinase C inhibitors were able to do so instead. They required to be vitamin D-rich and have VDR, showing that the fundamental calcium transport mechanism remained intact. L type channel activators, such as the BAY K-8644 and the phorbol esters, might stimulate transcaltachia in a manner similar to that of the 1,25(OH)<sub>2</sub>D, on the other hand.

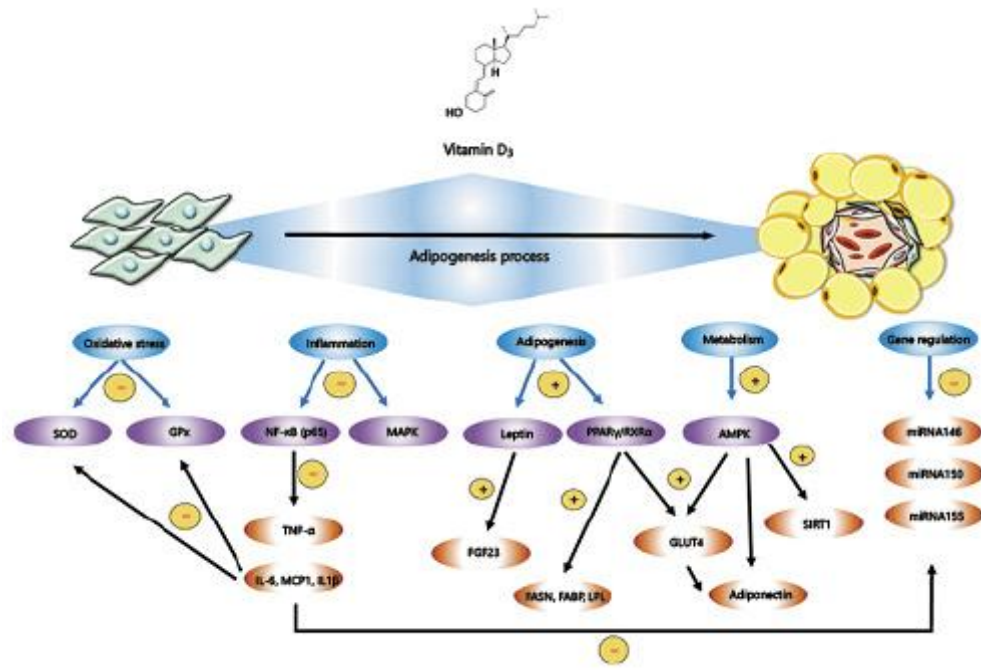
### **VITD AND ADIPOGENESIS**

“The effects of 1,25(OH)<sub>2</sub>-D are still unclear in the adipogenesis process. Adipogenesis is the process of cell differentiation by which preadipocytes become mature adipocytes. In human cells, 1,25(OH)<sub>2</sub>-D stimulates adipogenesis by the upregulation of gene expression enzymes of the lipogenesis process such as fatty acid synthase (FASN), fatty acid binding protein (FABP), and peroxisome proliferator activator receptor (PPAR)- $\gamma$ , which is the main transcription factor involved in the adipogenic differentiation. However, 1,25(OH)<sub>2</sub>-D inhibits this process in mice 3T3-L1 pre-adipocytes, by downregulating the transcription factors C/EBP $\alpha$ , C/EBP $\beta$ , and PPAR- $\gamma$ , and sequestering the nuclear receptor retinoic X receptor (RXR), a member of the nuclear receptor superfamily. On the contrary, 1,25(OH)<sub>2</sub>-D up-regulates FASN and lipoprotein

lipase (LPL) in human subcutaneous preadipocytes, a process that might be mediated by an increased expression of PPAR- $\gamma$ .

Generally, vitD, both 25(OH)-D and 1,25(OH)<sub>2</sub>-D, are capable of promoting adipogenic differentiation into mature adipocytes due to the presence of 1 $\alpha$ -hydroxylase in mature adipocytes. Additionally, 1,25(OH)<sub>2</sub>-D stimulates the translocation of the glucose transporter 4 (GLUT4) into the membrane, promotes adiponectin secretion and the expression of typical adipocyte genes, such as leptin, and inhibits the expression of uncoupling proteins in vitro. Thus, VDR directly inhibits the expression of the uncoupling protein-1 (UCP1), the critical protein for uncoupling fatty acid oxidation in brown AT (BAT). As a matter of fact, this process occurs cell autonomously and is independent of the physiologic VDR hormone ligand, 1,25(OH)<sub>2</sub>-D. On the other hand, leptin is capable of increasing the secretion of fibroblast growth factor 23, which is a negative regulator of renal 1 $\alpha$ -hydroxylase, thereby closing a negative feedback loop. However, an inhibitory effect of 1,25(OH)<sub>2</sub>-D on leptin secretion by human adipocytes has been observed in vitro. Indeed, the effect of vitD supplementation on leptin levels in humans remains poorly investigated and should be further addressed.

Vitamin D effects on adipogenesis and inflammation. AMPK, adenosine monophosphate kinase; FABP4, fatty acid-binding protein 4; FASN, fatty acid synthase; FGF23, fibroblast growth factor 23; GLUT4, glucose transporter 4; GPX, glutathion peroxidase; IL1 $\beta$ , interleukin 1-beta; IL6, interleukin 6; LPL, lipoprotein lipase; MAPK, mitogen-activated protein kinase; MCP1, monocyte chemotactic protein 1; NF- $\kappa$ B, nuclear factor kappa-light-chain-enhancer of activated B cells; PPAR $\gamma$ , peroxisome proliferator-activator receptors gamma; RXR $\alpha$ , retinoid X receptor; SIRT1, sirtuin 1; SOD, superoxide dismutase; TNF $\alpha$ , tumor necrosis factor alpha.”



**Figure 1**vitD main effects on adipogenesis in human adipocytes.

“VitD regulates the adipokine secretion in adipocytes, such as adiponectin, leptin, and resistin. Adiponectin is an anti-inflammatory and insulin-sensitizing hormone, which is the major adipokine secreted by adipocytes. VitD is associated with low levels of adiponectin in children with obesity, and vitD supplementation ameliorates systemic inflammatory biomarkers, including adiponectin, in the subjects with type 2 diabetes. However, no effect of 1,25(OH)2-D on adiponectin expression in human adipocyte culture has been observed.”

- **VitD and Inflammation**

Obesity is significantly linked to low-grade inflammation and the synthesis and release of proinflammatory mediators. ' As a result, immune cells play a critical role in maintaining normal AT and immunological homeostasis in humans. In vitro studies have shown that 1,25(OH)2D has an anti-inflammatory effect on adipocytes, which is linked to obesity. Adipocytes emit less chemokines and cytokines, and monocytes are less likely to chemotact. The cytokine release from visceral AT (VAT) was reduced by vitD in AT explants, but not from SAT, which was unaffected. A decrease in adenosine monophosphate-activated protein kinase (AMPK) in the VAT of obese persons has been shown to be strongly linked to inflammation in the AT. Another factor that may help prevent obesity and obesity-related metabolic dysfunction is AMPK's ability to boost sirtuin 1 by raising NAD/NADH ratio and decreasing AT macrophage infiltration and

inflammation. Oxidation-related gene mRNA levels were reduced by vitD deficiency, according to a recent research. It was also shown that in obese rats, sirtuin 1 and AMPK activity was significantly reduced. Due to the detrimental effects of vitD shortage on AT growth, immune cell infiltration, and inflammatory condition, it seems that vitD plays a favourable function in adipocyte metabolic metabolism and obesity development.

The in vitro regulation of miR expression in adipocytes by vitD was recently shown. When 1,25(OH)<sub>2</sub>-D was preincubated with human adipocytes in the presence of tumour necrosis factor (TNF)-, the expression of three miRNAs (miR-146a; miR-150; and miR-155) was severely inhibited. Additionally, the three miRs were shown to be upregulated in mice given a high-fat diet supplemented with vitD or an epididymal white AT diet heavy in fat. As a result, in vitro and in vivo studies utilising aP2-p65 transgenic mice demonstrated that NF-B signalling is necessary for the induction of these miRs. Anti-NF-B signalling was detected through suppression of I-B and I-B in mouse adipocytes, and this might be a driving molecular mechanisms (Fig. 1).

In addition to the reported benefits, vitD lowers levels of proinflammatory cytokines and chemokines in mice following injections of lipopolysaccharide and in obese animals created by a high-fat diet. In normal-weight participants, there was a negative correlation between serum 25(OH)D concentration and plasma interleukin (IL)-6 and TNF- levels. Despite this, neither serum nor AT inflammatory indicators were altered by vitD in obese participants.

The NF-B and mitogen-activated protein kinase signalling pathways are inhibited by VitD, which has anti-inflammatory properties. In addition, vitD lowered the expression of toll-like receptors, which is elevated in immune cells and AT from obese persons alike. A transmembrane protein, the toll-like receptor (TLR), triggers conventional signalling cascades that lead to NF-B activation as well as the generation of inflammatory cytokines, such as TNF-. Human adipocytes incubated with 1,25(OH)<sub>2</sub>-D show a reduction in inflammatory markers such IL-6, MCP-1 and IFN- (mRNA and protein level). The TNF—mediated pro-inflammatory marker is also downregulated by this therapy. 1,25(OH)<sub>2</sub>-D has been shown to have a comparable impact on the expression of proinflammatory markers in adipocyte-macrophage co-culture systems. In this experiment, the role of VDR and NF-B was validated. Last but not least, 1,25(OH)<sub>2</sub>-D treatment causes p38 to be dephosphorylated, which has been associated to the transcriptional activation of several Dusp family members. As a result, increased glucose absorption and phosphorylation of

AKT were found, suggesting that vitD insufficiency may be associated to low-grade inflammation. Vitamin D supplementation has been shown to reduce inflammation and oxidative stress in obese rats fed a high-fat diet. As a result, both in normal-weight and obese rats fed a high-fat diet, vitD therapy significantly reduced AT TNF- concentrations. vitD-supplemented high-fat diet-fed AT had considerably higher amounts of oxidative stress indicators like as superoxide dismutase and glutathione peroxidase, as well as lower levels of monocyte chemotactic protein-1 (Fig. 1). In obese rats, vitD reduced AT oxidative stress and inflammation. As a result, vitD has a significant impact on adipocyte and AT inflammation because of its capacity to reduce phosphorylation and nuclear translocation of NF-B p65 and, hence, proinflammatory and oxidative stress indicators. A lot more research is required to figure out exactly how this works.

According to the wide scientific literature, there is a definite correlation between BMI and vitD insufficiency in both adults and children. A causal link between elevated levels of serum vitD and obesity will need to be established in future prospective research. Vitamin D supplementation has a lot less research available, and it's not obvious whether it improves obesity's metabolic profile. To better understand how vitD supplementation affects obesity-related biomarkers, future randomised controlled trials should concentrate on refining the research design, especially for studies evaluating vitD, as most trials contained two or more trial treatments (i.e., weight loss, resistance or exercise training interventions, with oral vitD supplementation). For one thing, the studies had a limited sample size and varied doses and duration of vitamin D. In future research, longer treatment durations with clinically safe greater dosages may be warranted.

It seems that genetic factors have a role in determining 25(OH)D levels in the bloodstream. The examination of genes associated with vitamin D in obese individuals is also an intriguing prospect. Some research, although not all, have shown an association between obesity and VDR genetic variations. In addition, there has been no apparent consensus in the results across research for any specific demographic. More research is needed to determine whether or not VDR genetic variants contribute significantly to the development of obesity-related symptoms. This conclusion is supported by an overwhelming number of studies that have found no association between VDR genetic variants and obesity symptoms, as well as studies that have found conflicting results for specific VDR SNPs. To wrap things up, while the majority of the

studies with null findings failed to get significant P-values following multiple testing correction, it may be beneficial to conduct research with bigger and more homogeneous populations. The "plausible" notion that VDR is a key obesity gene may be supported if research reach agreement in demonstrating repeatable relationships in this manner.

## **CONCLUSION**

When it comes to calcium homeostasis and bone mineralization, vitamin D has long been regarded as an essential nutrient (Reichel et al,1989). It has been shown that vitamin D has a significant impact on the function of muscles (Ceglia et al,2013). There is a favourable link between 25(OH)D3 levels and physical performance in the elderly, as well as a negative correlation with the risk of falls. The structure and function of skeletal muscle may be affected by vitamin D, which is well-known. A lack of vitamin D is associated with a decrease in muscle mass and strength, regardless of age. Vitamin D deficiency has been reported to cause mostly type II muscle fibre atrophy in skeletal muscle biopsy samples. Vitamin D supplementation has been found in human trials to prevent muscular atrophy. Systemic illnesses such as autoimmune disorders, diabetes, cancer, and AIDS are all associated with skeletal muscle wasting/atrophy. Protein degradation and synthesis pathways in the muscle are imbalanced, resulting in muscular atrophy. There are three primary proteolytic systems in the skeletal muscle, which all play a role in protein breakdown. Three pathways are involved: the ubiquitin-proteasome, the lysosomal, and the calpain. In both animals and humans, a lack of vitamin D has been linked to muscle atrophy. Vitamin D insufficiency causes muscle atrophy, however no research have examined the role of these three proteolytic processes.

Studies have shown that oxidative stress may cause muscular atrophy under different physiological situations. As a result, it's unclear if oxidative stress plays a role in muscle atrophy or whether it is a result of it. Vitamin D is said to protect cells from the harmful effects of free radicals by acting as an antioxidant. Vitamin D deficiency-induced muscle loss has not been linked to oxidative stress, at least not in studies that have been published yet. No studies have shown that vitamin D can reverse the proteolytic alterations caused by oxidative stress, or that it can act as an antioxidant or a protective agent in muscle. In the adipose tissue, there is a vitamin D receptor and the enzymes necessary to convert vitamin D into the active hormone form. Cell culture models used for these investigations showed that 1,25(OH)2D3 may have an impact on lipid metabolism and storage. Vitamin D/VDR in vivo research on obesity are few and far

between. A lean phenotype and resilience to obesity-inducing high-fat diets are linked to a lack of VDR, according to one research. Another research found that adipogenesis was inhibited when VDR was downregulated (Blumberg et al.2006). There are no reports on the effect of vitamin D deficiency on body fat distribution in rats fed a diet lacking in vitamin D.

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